SHORT COMMUNICATION

Arabinoxylan-oligosaccharides (AXOS) reduce preneoplastic lesions in the colon of rats treated with 1,2-dimethylhydrazine (DMH)

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Abstract

Background Prebiotics are non-digestible compounds that beneficially affect the host by stimulating the growth and/or activity of one or a limited number of resident colonic bacteria in the gut. Reported beneficial effects of prebiotics include reduced gut infections, better absorption of minerals, and notably, antitumorigenic effects. Arabinoxylan (AX)-oligosaccharides (AXOS) have been suggested to exert prebiotic effects in the gut, but their effect on colon carcinogenesis has not been studied so far.

Aim of the study To test the effect of AXOS in a rat colon carcinogenesis model.

Methods We determined the occurrence of two types of preneoplastic lesions [aberrant crypt foci (ACF) and mucin depleted foci (MDF)] in the colon of rats treated with the colon carcinogen 1,2-dimethylhydrazine (DMH) and fed either a control diet or a diet containing AXOS (4.8% w/w) (15 rats in each group).

Results Thirteen weeks after DMH treatment, MDF counts were significantly lower in the entire colon of AXOS fed rats (MDF/colon were 7.5 ± 0.6 and 5.5 ± 0.6 ,

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W. F. Broekaert · J. A. Delcour · C. M. Courtin Laboratory of Food Chemistry and Biochemistry and Leuven Food Science and Nutrition Research Centre (LFoRCe), Katholieke Universiteit Leuven, Kasteelpark Arenberg 20 bus 2463, 3001 Leuven, Belgium in Control and AXOS groups, respectively, means \pm SE, P < 0.05). Although the number of ACF in the entire colon was not significantly different between Control and AXOS fed rats, AXOS fed rats had significantly fewer ACF in the distal part of the colon than Control group rats (ACF/distal colon were 135.5 ± 15 and 84.4 ± 11 , in Control and AXOS groups, respectively, means \pm SE, P < 0.05). Conclusions The present study shows that dietary intake of AXOS by rats reduces the occurrence of two types of preneoplastic lesions, thus suggesting a chemopreventive effect on colon carcinogenesis that should be confirmed in a long-term carcinogenesis experiment.

Keywords Colon carcinogenesis · Arabinoxylan-oligosaccharides · Preneoplastic lesions

Introduction

Arabinoxylan (AX), formerly also referred to as pentosan, is a major constituent of cereal cell walls and consists of a backbone of β -D-xylopyranosyl residues (xylose), some of which are mono- or disubstituted with α -L-arabinofuranosyl residues (arabinose) [1, 9, 25]. From a dietary point of view, AX has raised interest as some types of oligosaccharides derived from it by enzymatic or chemical cleavage have antioxidant properties, due to the presence of hydroxycinnamic acids, mainly ferulic acid linked through ester bonds to the arabinose residues of AX [1, 26, 30]. Moreover, arabinoxylan-oligosaccharides (AXOS) have been suggested to exert a prebiotic effect in the gut [10, 12, 22, 41]. Prebiotics are non-digestible compounds that beneficially affect the host by stimulating the growth and/ or activity of one or a limited number of resident colonic bacteria in the gut [21, 38]. Among the beneficial effects



of prebiotics, particular attention has been paid to their purported effect on colon cancer. Several experimental studies indicate in fact that prebiotics, such as fructooligosaccharides (FOS), xylooligosaccharides (XOS), galactooligosaccharides (GOS) are protective against colon carcinogenesis [17, 29, 34, 35, 38–40]. However, the effect of AXOS on colon carcinogenesis has not been studied so far. Therefore, in the present study we tested the effect of AXOS on colon carcinogenesis determining preneoplastic lesions [aberrant crypt foci (ACF) and mucin depleted foci (MDF)] [7, 11, 19] in rats treated with the colon carcinogen 1,2-dimethylhydrazine (DMH), a widely used experimental model to test the potential chemopreventive activity of food components [11].

Materials and methods

Materials

1,2-dimethylhydrazine (DMH) was purchased from Sigma (Milan, Italy). Dietary components for the preparation of the AIN76 diet (see below) were purchased from Piccioni (Gessate, Milan, Italy). The AXOS preparation used in this study was obtained by xylanolytic hydrolysis of wheat bran, as described by Swennen et al. [36] and was provided by Fugeia NV (Leuven, Belgium). Table 1 presents the characteristics of the AXOS preparation, which had an average degree of polymerization (avDP) of five and an average degree of arabinose substitution (avDAS) of 0.21. Total, reducing end, and free saccharide contents were determined by gas chromatographic analysis [13] and calculation of AXOS content, avDP and avDAS was done as described earlier [13], except that no correction for arabinogalactan content was made due to low galactose content. Moisture and ash contents of the preparation were analyzed according to AACCI methods 44-19 and 08-01, respectively [2]. Protein content was determined according to a Dumas combustion method, using an automated Dumas protein analysis system (EAS varioMax N/CN, Elt, Gouda, The Netherlands), an adaptation of the AOAC Official Method for protein determination [3] and using 6.25 as the nitrogen protein conversion factor. Ferulic acid (FA) content was determined by High Performance Liquid Chromatography with UV light detection at 310 nm according to Hartmann et al. [23], using o-coumaric acid as an internal standard. Total FA content is determined after saponification of the sample in 2 M NaOH for 18 h at room temperature under nitrogen atmosphere. Free FA is determined without prior saponification. Glucuronic acid (GluA) content was determined colorimetrically by the m-phenylphenol method using D-glucuronic acid for calibration [4].



Table 1 Characterization of the AXOS preparation

Compositional parameter	AXOS preparation
Moisture (%)	3.0
$AXOS^a$ (% dm) b	83.4
avDAS ^c of AXOS: 0.21	
avDP ^d of AXOS: 5	
Ferulic acid bound to AXOSe (% dm)	1.5
Glucuronic acid bound to AXOSf (% dm)	1.0
Glucose as part of poly/oligosaccharides ^g (% dm)	12.4
Galactose as part of poly/oligosaccharides ^g (% dm)	0.6
Mannose as part of poly/oligosaccharides ^g (% dm)	0.2
Free monosaccharidesh (% dm)	1.6
Protein	0.4
Ash (% dm)	0.5

- a AXOS content = 0.88 \times [arabinose_{TOT} arabinose_{FREE}] + [xylose_{RED} xylose_{FREE}] + 0.88 \times [xylose_{TOT} xylose_{RED}]
- b dm = dry matter
- c avDAS = [arabinose_{TOT} arabinose_{FREE}]/[xylose_{TOT} xylose_{FREE}]
- d avDP = {[arabinose_{TOT} arabinose_{FREE}] + [xylose_{TOT} xylose_{FREE}]}/[xylose_{RED} xylose_{FREE}]
- $^{\rm e}~$ bound ferulic acid = 0.907 \times [ferulic acid $_{TOT}-$ ferulic acid $_{FREE}$]; free ferulic acid content <0.01% dm
- $^{\rm f}$ bound glucuronic acid = 0.907 \times [glucuronic acid $_{\rm TOT}$]; free glucuronic acid content <0.01% dm
- $^{\rm g}$ glucose content as part of poly/oligosaccharides = 0.9 × [glucose_{TOT} glucose_{FREE}] + [glucose_{RED} glucose_{FREE}]; mannose and galactose as part of poly/oligosaccharides are calculated in the same way. Total poly/oligosaccharides is the sum of AXOS, ferulic acid, and glucuronic acid bound to AXOS, glucose—, galactose— and mannose as part of poly/oligosaccharides
- $^{\rm h}$ free monosaccharides = glucose_{FREE} + xylose_{FREE} + arabinose FREE. Other free monosaccharides are <0.05% dm

Animals and treatments

We used 5–6-week-old male F344 rats (Nossan, Correzzana, Milan, Italy), which were randomly allocated to two groups (15 rats per group). The animals were housed in plastic cages with wire tops and maintained at a temperature of 22°C, with a 12:12-h light-dark cycle, according to the European Union Regulations on the Care and Use of Laboratory Animals [16]. The experimental protocol was approved by the Commission for Animal Experimentation of the Ministry of Health, Rome, Italy. A control group (Control) was fed a high-fat diet based on the AIN76 diet [18], but with a higher portion of lipids (230 g/kg corn oil) and a lower level of cellulose (20 g/kg). The source of carbohydrates in this diet was sucrose (341 g/kg), maize starch (60 g/kg), and maltodextrins (Maldex 150, Syral, 60 g/kg). A treatment group (AXOS) was fed the same diet

as the Control group, but this diet contained 60 g/kg of AXOS preparation instead of maltodextrins, providing a pure AXOS content of 48 g/kg. Ten days after the beginning of the experimental diets, both groups were treated twice, 1 week apart, with 100 mg/kg of DMH by subcutaneous injection. Dietary treatment (ad libitum for both groups) continued until the end of the experiment, i.e., 13 weeks after the first injection with DMH, when rats were sacrificed with CO₂ asphyxiation.

Determination of ACF and MDF

At sacrifice, the cecum and colon were dissected and washed with saline. The cecal contents and cecal walls were weighed. The colon was pinned flat on a polystyrene board to reduce any folding of the colon mucosa which would otherwise have interfered with a good visualization of MDF and fixed in buffered formalin [18]. The number of ACF and their multiplicity (number of aberrant crypts forming each ACF) were determined after methylene blue staining [7]. After ACF determination, methylene blue stained colons were kept in formalin and then processed with the high-iron diamine Alcian blue staining to determine the number of MDF and their multiplicity (number of crypts forming each MDF) [7]. Regarding the localization of ACF and MDF along the colon, the entire colon, extending from the anal to the cecal ends, was divided in two equally long parts: one proximal and one distal.

Statistical analysis

Data obtained from individual rats in the two groups were analyzed with two tailed t-test (unpaired samples) calculating the contrasts between means. Differences were considered statistically significant when P was <0.05.

Results

Body weight and cecal parameters

There were no differences in body weights between the two experimental groups. Body weight at the beginning of the experiment was 159 ± 2.4 and 158 ± 2.4 g in Control and AXOS groups, respectively, and 357 ± 6.5 and 350 ± 6.5 g at the end of the experiment (means \pm SE).

Cecal content at sacrifice was significantly higher in the AXOS than in the Control groups, while the weight of the cecal wall did not differ between groups (Table 2).

Table 2 Cecal parameters in rats treated with Control or AXOS diet

	Control	AXOS
Cecal content (wet weight, g)	1.9 ± 0.2	3.2 ± 0.3**
Cecal wall (wet weight, g)	1.2 ± 0.1	1.5 ± 0.1

Values represent means \pm SE n=12, since this determination was carried out in a random subgroup of rats for each group **Significantly different when compared with Control group (P < 0.001)

Preneoplastic lesions: aberrant crypt foci (ACF) and mucin depleted foci (MDF) determination

The number of ACF determined in the entire colon of rats was not different between Control and AXOS fed rats (Fig. 1, panel a). Similarly, the multiplicity of these lesions, i.e., the number of crypts forming each ACF, did not vary (crypts/ACF were 2.6 ± 0.1 and 2.7 ± 0.1 in Control and AXOS groups, respectively, means \pm SE). Interestingly, when considering the distribution of ACF along the colon, we observed that in the distal part of the colon, the number of ACF in the AXOS group was significantly lower than that in the Control group (Fig. 1, panel b). No difference between the two groups was observed for the proximal part of the colon.

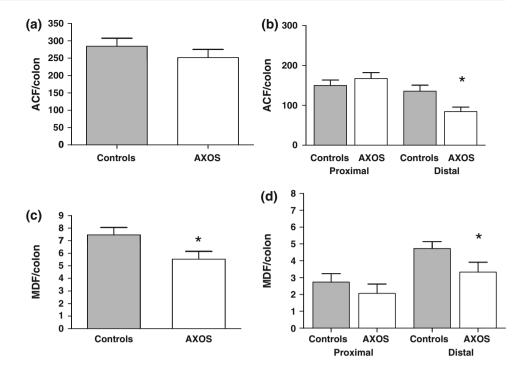
The number of MDF in the entire colon was significantly lower in the animals fed AXOS than in the Control group (Fig. 1, panel c). The multiplicity of MDF was slightly lower in the AXOS group, but this effect did not attain statistical significance (crypts/MDF 13.2 ± 4.2 and 8.7 ± 2.2 in the Control and AXOS fed groups, respectively, means \pm SE). Considering the distribution of MDF along the colon in the two groups, we also found, as observed for ACF, a statistically significant effect in the distal part of the colon (Fig. 1, panel d).

Discussion

In the gut, prebiotics cause changes in the intestinal microbiota which have been associated with improved overall health, reduced gut infections, and better absorption of minerals [21, 29, 37]. Moreover, the results of several studies, mainly in rodents, pointed out that prebiotic compounds such as GOS, XOS, FOS, or inulin may be protective against colon carcinogenesis [17, 29, 34, 35, 38–40]. For instance, FOS and inulin have been reported to reduce ACF in rats [34, 35]; similarly, it has been reported that synergy-1, an inulin-derived prebiotic compound, significantly reduced colon cancer in carcinogen-treated rats [17]. Other prebiotics such as GOS have also been shown to reduce colon carcinogenesis, with only slight effects on preneoplastic lesions [40]. Arabinoxylan-oligosaccharides



Fig. 1 Number of ACF and MDF in the colon of Control (gray bars) or AXOS (white bars) group rats. Panels $\bf a$ and $\bf c$: number of ACF and MDF, respectively, in the entire colon; panels $\bf b$ and $\bf d$: number of ACF and MDF, respectively, in the proximal and distal colon of the two experimental groups. *Significantly different (P < 0.05) when compared with Control group. Bars are means + SE, n = 15



(AXOS) have been suggested to exert prebiotic effects in various experimental settings [13-15, 36, 37]. In rats, AXOS with an average degree of polymerization of five, as in the present study, increased acetate and butyrate concentrations in the colon, reduced intestinal protein fermentation, and increased concentrations of cecal bifidobacteria [37]. In chickens, AXOS increased cecal bifidobacteria while reducing potentially pathogenic Salmonella strains [14, 15, 41]. Moreover, addition of AXOS to the diet of healthy human volunteers caused a significant reduction of urinary ammonia excretion and higher excretion via the feces, thus suggesting beneficial effects [10]. Given the earlier reported beneficial effects of AXOS, we were interested in studying their possible protective effect on colon carcinogenesis. Although tumors are the best endpoints for evaluation of the chemopreventive effects of dietary agents, preneoplastic lesions such as aberrant crypt foci (ACF) and mucin depleted foci (MDF) can be also used as cancer endpoints avoiding long-term carcinogenesis experiments, which are time and animal consuming [7, 11, 31–33]. The results of the present study clearly show a significant effect of AXOS in reducing the number of MDF, early precancerous lesions of colon carcinogenesis [7, 20]. The effect of AXOS on MDF was evident in the entire colon, with marked effects for the distal part. Moreover, the MDF in the AXOS group tended to be smaller, as demonstrated by the lower multiplicity, but this effect did not attain statistical significance. Contrary to the clear-cut effect on MDF, the results on ACF showed that, while the number of ACF in the entire colon was not

impacted by AXOS, there was a statistically significant reduction in the distal part of the colon. Using an in vitro simulator of the human intestinal microbial ecosystem it has been demonstrated that AXOS fermentation in the colon occurs more distally than fermentation of the well studied prebiotic inulin [22], a phenomenon which may explain why the suppressive effect of AXOS on ACF and MDF is more pronounced in the distal than in the proximal colon of DMH treated rats. Although several studies showed that prebiotics have chemopreventive activity in colon carcinogenesis [17, 29, 34, 35, 38–40], only a few investigated the effect of these compounds along the various parts of the colon, thus making difficult to compare our results with other studies. For instance, it has been reported that inulin reduced ACF/colon in both proximal and distal colon of rats and mice [39], while FOS reduced ACF in mice only in the distal colon [6]. Moreover in a study by Hsu et al. [24], in which XOS, structurally similar to AXOS, and FOS reduced ACF, the enumeration of these lesions was carried out only in the distal part of the colon, while no data were presented for either the proximal or the entire colon. It is also interesting to note that the effect of dietary variations on colon cancer has been suggested to be more evident in the distal than in the proximal colon [27, 28].

Addition of AXOS to the diet did not affect the body weight of the animals, consistent with an earlier study [37]. The marked increase in cecal content caused by AXOS addition to the diet has been observed before for other prebiotics such as XOS, FOS, and inulin [8, 24, 42], and is



considered to be a normal physiological response to the presence of high concentrations of fermentable non-digestible oligosaccharides in the diet [5].

In conclusion, we showed a protective effect of AXOS on two types of preneoplastic lesions in a rat colon carcinogenesis model. We believe that this result is indicative of a possible chemopreventive effect of AXOS on colon carcinogenesis that should be confirmed in a long-term carcinogenesis experiment.

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Conflict of interests statement WFB, CMC, IEJAF, and JAD hold stock (options) of Fugeia NV. WFB and JAD are directors of Fugeia NV.

References

- Andersson R, Aman P (2001) Cereal arabinoxylan: occurrence, structure and properties. In: McCleary BV, Prosky L (eds) Advanced dietary fibre technology. Blackwell Science Ltd, Oxford, pp 301–314
- Anonymous (2000) Approved methods of the American Association of Cereal Chemists, 10th edn. AACC, St. Paul, MN, USA
- AOAC (1995) Official methods of analysis, 16th edition. Method 990.03. Association of Official Analytical Chemists, WA, USA
- Blumenkrantz N, Asboe-Hansen G (1973) New method for quantitative determination of uronic acids. Anal Biochem 54: 484–489
- Boyle FG, Wrenn JM, Marsh BB, Anderson WI, Angelosanto FA, McCartney AL, Lien EL (2008) Safety evaluation of oligofructose: 13 week rat study and in vitro mutagenicity. Food Chem Toxicol 46:3132–3139
- Buddington KK, Donahoo JB, Buddington RK (2002) Dietary oligofructose and inulin protect mice from enteric and systemic pathogens and tumor inducers. J Nutr 132:472–477
- Caderni G, Femia AP, Giannini A, Favuzza A, Luceri C, Salvadori M, Dolara P (2003) Identification of mucin-depleted foci in the unsectioned colon of azoxymethane-treated rats: correlation with carcinogenesis. Cancer Res 63:2388–2392
- Campbell JM, Fahey GC Jr, Wolf BW (1997) Selected indigestible oligosaccharides affect large bowel mass, cecal and fecal short-chain fatty acids, pH and microflora in rats. J Nutr 127:130–136
- Cleemput G, Roels SP, van Oort MG, Grobet PJ, Delcour JA (1993) Heterogeneity in the structure of water soluble arabinoxylans in European wheat flours of variable breadmaking quality. Cereal Chem 70:324–329
- Cloetens L, De Preter V, Swennen K, Broekaert WF, Courtin CM, Delcour JA, Rutgeerts P, Verbeke K (2008) Dose-response effect of arabinoxylo-oligosaccharides on gastrointestinal motility and on colonic bacterial metabolism in healthy volunteers. J Am Coll Nutr 27:512–518
- Corpet DE, Pierre F (2003) Point: from animal models to prevention of colon cancer. Systematic review of chemoprevention in min mice and choice of the model system. Cancer Epidemiol Biomarkers Prev 12:391–400
- Courtin CM, Swennen K, Broekaert WF, Swennen Q, Buyse J, Decuypere E, Michiels CW, De Ketelaere B, Delcour JA (2008)

- Effects of dietary inclusion of xylooligosaccharides, arabinoxylooligosaccharides and soluble arabinoxylan on the microbial composition of caecal contents of chickens. J Sci Food Agric 88:2517–2522
- Courtin CM, Broekaert WF, Aerts G, Van Craeyveld V, Delcour JA (2009) Occurrence of arabinoxylo-oligosaccharides and arabinogalactan peptides in beer. J Am Soc Brew Chem 67: 112–117
- 14. Courtin CM, Swennen K, Broekaert WF, Lescroart O, Onagbesan O, Buyse J, Decuypere E, Van de Wiele T, Marzoratti M, Verstraete W, Huyghebaert G, Delcour JA (2008) Dietary inclusion of wheat bran arabinoxylooligosaccharides has beneficial nutritional effects on chickens. Cereal Chem 85:607–613
- Eeckhaut V, Van Immerseel F, Dewulf J, Pasmans F, Haesebrouck F, Ducatelle R, Courtin CM, Delcour JA, Broekaert WF (2008) Arabinoxylooligosaccharides from wheat bran inhibit Salmonella colonization in broiler chickens. Poult Sci 87: 2329–2334
- European Community: European Community Regulations on the Care and Use of Laboratory Animals (1986) (Law 86/609/EC)
- 17. Femia AP, Luceri C, Dolara P, Giannini A, Biggeri A, Salvadori M, Clune Y, Collins KJ, Paglierani M, Caderni G (2002) Antitumorigenic activity of the prebiotic inulin enriched with oligofructose in combination with the probiotics Lactobacillus rhamnosus and Bifidobacterium lactis on azoxymethane-induced colon carcinogenesis in rats. Carcinogenesis 23:1953–1960
- Femia AP, Caderni G, Bottini C, Salvadori M, Dolara P, Tessitore L (2007) Mucin-depleted foci are modulated by dietary treatments and show deregulation of proliferative activity in carcinogen-treated rodents. Int J Cancer 120:2301–2305
- Femia AP, Dolara P, Caderni G (2004) Mucin-depleted foci (MDF) in the colon of rats treated with azoxymethane (AOM) are useful biomarkers for colon carcinogenesis. Carcinogenesis 25:277–281
- Femia AP, Giannini A, Fazi M, Tarquini E, Salvadori M, Roncucci L, Tonelli F, Dolara P, Caderni G (2008) Identification of mucin depleted foci in the human colon. Cancer Prev Res 1:562–567
- Gibson GR, Roberfroid MB (1995) Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. J Nutr 125:1401–1412
- 22. Grootaert C, Van den Abbeele P, Marzorati M, Broekaert WF, Courtin CM, Delcour JA, Verstraete W, Van de Wiele T (2009) Comparison of prebiotic effects of arabinoxylan oligosaccharides and inulin in a simulator of the human intestinal microbial ecosystem. FEMS Microbiol Ecol 69:231–242
- Hartmann G, Piber M, Koehler P (2005) Isolation and chemical characterisation of water-extractable arabinoxylans from wheat and rye during breadmaking. Eur Food Res Technol 221:487–492
- Hsu CK, Liao JW, Chung YC, Hsieh CP, Chan YC (2004) Xylooligosaccharides and fructooligosaccharides affect the intestinal microbiota and precancerous colonic lesions development in rats. J Nutr 134:1523–1528
- Izydorczyk MS, Biliaderis CG (1995) Cereal arabinoxylans: in structure and physicochemical properties. Carbohydr Polym 28:33–48
- Katapodis P, Vardakou M, Kalogeris E, Kekos D, Macris BJ, Christakopoulos P (2003) Enzymic production of a feruloylated oligosaccharide with antioxidant activity from wheat flour arabinoxylan. Eur J Nutr 42:55–60
- 27. Kimura Y, Kono S, Toyomura K, Nagano J, Mizoue T, Moore MA, Mibu R, Tanaka M, Kakeji Y, Maehara Y, Okamura T, Ikejiri K, Futami K, Yasunami Y, Maekawa T, Takenaka K, Ichimiya H, Imaizumi N (2007) Meat, fish and fat intake in relation to subsite-specific risk of colorectal cancer: The Fukuoka Colorectal Cancer Study. Cancer Sci 98:590–597



- 28. Koushik A, Hunter DJ, Spiegelman D, Beeson WL, van den Brandt PA, Buring JE, Calle EE, Cho E, Fraser GE, Freudenheim JL, Fuchs CS, Giovannucci EL, Goldbohm RA, Harnack L, Jacobs DR Jr, Kato I, Krogh V, Larsson SC, Leitzmann MF, Marshall JR, McCullough ML, Miller AB, Pietinen P, Rohan TE, Schatzkin A, Sieri S, Virtanen MJ, Wolk A, Zeleniuch-Jacquotte A, Zhang SM, Smith-Warner SA (2007) Fruits, vegetables, and colon cancer risk in a pooled analysis of 14 cohort studies. J Natl Cancer Inst 99(19):1471–1483
- Macfarlane GT, Steed H, Macfarlane S (2008) Bacterial metabolism and health-related effects of galacto-oligosaccharides and other prebiotics. J Appl Microbiol 104:305–344
- Ohta T, Semboku N, Kuchii A, Egashira Y, Sanada H (1997) Antioxidant activity of corn bran cell-wall fragments in the LDL oxidation system. J Agric Food Chem 45:1644–1648
- Pierre F, Santarelli R, Taché S, Guéraud F, Corpet DE (2008) Beef meat promotion of dimethylhydrazine-induced colorectal carcinogenesis biomarkers is suppressed by dietary calcium. Br J Nutr 99:1000–1006
- 32. Plate AY, Gallaher DD (2006) Effects of indole-3-carbinol and phenethyl isothiocyanate on colon carcinogenesis induced by azoxymethane in rats. Carcinogenesis 27:287–292
- Pretlow TP, Pretlow TG (2005) Mutant KRAS in aberrant crypt foci (ACF): initiation of colorectal cancer? Biochim Biophys Acta 1756:83–96
- Reddy BS, Hamid R, Rao CV (1997) Effect of dietary oligofructose and inulin on colonic preneoplastic aberrant crypt foci inhibition. Carcinogenesis 18:1371–1374
- 35. Rowland IR, Rumney CJ, Coutts JT, Lievense LC (1998) Effect of bifidobacterium longum and inulin on gut bacterial metabolism

- and carcinogen-induced aberrant crypt foci in rats. Carcinogenesis 19:281–285
- Swennen K, Courtin CM, Lindemans GCJE, Delcour JA (2006)
 Large scale production and characterisation of wheat bran arabinoxylooligosaccharides. J Sci Food Agric 86:1722–1731
- 37. Van Craeyveld V, Swennen K, Dornez E, Van de Wiele T, Marzorati M, Verstraete W, Delaedt Y, Onagbesan O, Decuypere E, Buyse J, De Ketelaere B, Broekaert WF, Delcour JA, Courtin CM (2008) Structurally different wheat-derived arabinoxylooligosaccharides have different prebiotic and fermentation properties in rats. J Nutr 138:2348–2355
- Van Loo J (2004) Prebiotics promote good health. The basis, the potential, and the emerging evidence. J Clin Gastroenterol 38:S70–S75
- Verghese M, Rao DR, Chawan CB, Shackelford L (2002) Dietary inulin suppresses azoxymethane-induced preneoplastic aberrant crypt foci in mature Fisher 344 rats. J Nutr 132:2804–2808
- Wijnands MV, Schoterman HC, Bruijntjes JB, Hollanders VM, Woutersen RA (2001) Effect of dietary galacto-oligosaccharides on azoxymethane-induced aberrant crypt foci and colorectal cancer in Fischer 344 rats. Carcinogenesis 22:127–132
- Yamada H, Itoh K, Morishita Y, Taniguchi H (1993) Structure and properties of oligosaccharides from wheat bran. Cereal Food World 38:490–492
- Younes H, Garleb K, Behr S, Rémésy C, Demigné C (1995) Fermentable fibers or oligosaccharides reduce urinary nitrogen excretion by increasing urea disposal in the rat cecum. J Nutr 125:1010–1016

